

Why Use TDM and Dashboards for Monoclonal Antibodies? The Promise of Individualized Therapy

Diane R Mould, PhD, FCP, FAAPS

Projections Research Inc.

USA

Scope of the lecture:

Therapeutic monoclonal antibodies (MAbs) have emerged as a major component of treatment for a wide variety of diseases including inflammatory diseases and oncology. With generally better safety profiles than chemical agents, MAbs have also demonstrated therapeutic failures that in some cases appear to be due to pharmacokinetic (PK) variability. This lecture will cover the primary factors that are predictive of between-patient variability for MAb PK, and will look at efforts to improve outcomes using therapeutic drug monitoring (TDM). This lecture will also introduce the concept of dashboard based dosing, which is a way of individualizing therapy based on the patients specific PK behavior and the promise of MAb therapy when TDM and dashboard guided individualized therapy is implemented.

Learning objectives:

1. Understand the pharmacokinetics of MAbs and factors that are commonly associated with MAb PK variability
2. Learn the relationships between MAb exposure and response and possible causes for therapeutic failure
3. Learn about the concept of dashboard guided dosing in conjunction with therapeutic drug monitoring

Extended abstract:

Background: Monoclonal antibodies (MAbs) exhibit complex pharmacokinetics (PK). Many factors impact anti-TNF MAb PK, altering MAb clearance and therefore the half-life: albumin, weight (particularly obesity), disease (severity, stage, co-morbidities), and concomitant administration of immunosuppressants (e.g. methotrexate). In addition, the pharmacodynamics or response to treatment can also impact MAb PK, and anti-drug antibodies (ADA) can be considered a pharmacodynamic response. These factors can alter MAb exposure, impacting on the likelihood of clinical response or subsequent loss of response (LOR) following an initial response. A potential cause of therapeutic failure and LOR is between-subject variability in exposure, which can arise from several sources, particularly MAb clearance and, for subcutaneously administered MAbs, from variability in the extent of absorption.

Clearance describes how a drug is removed from the body; and is inversely related to half-life with slower clearance resulting in longer half-life. Clearance determines trough concentrations – higher clearance is associated with lower trough levels for the same dose interval. Unlike small molecule drugs, which are often cleared by cytochrome P450 enzymes, MAbs are primarily cleared through proteolysis, although specific sites of catabolism have not been identified [1]. Proteolytic clearance is generally related to patient weight, with higher weight subjects having more rapid proteolytic clearance [2, 3]. Renal elimination generally does not contribute to MAb clearance, owing to their high molecular weight which limits glomerular filtration [4] although patients with focal segmental glomerulosclerosis (FSGS) have been found to have increased MAb clearance [5], which is a form of atypical clearance. Atypical clearance is often associated disease type and severity resulting from altered catabolic pathways or organ function. For example Beeken et al [6] and later, Kaplan et al [7] reported increased intestinal IgG clearance in patients with inflammatory bowel disease (IBD) that correlated with lesion severity. Similarly, Brandse [8] reported infliximab concentrations in feces from severe IBD patients that were associated poor response. Protein losing enteropathy was also noted in patients with systemic lupus erythematosus [9].

Target mediated clearance (TMC) is a saturable route of elimination that can be a substantial component of MAb clearance. The fraction of TMC to overall clearance depends on MAb concentration and target antigen expression, including MAb-antigen internalization and receptor turnover rate [10], which can result in both nonlinear and time dependent changes in clearance [11] and can also result in disease related differences in MAb PK. Antigens on cell surfaces may be shed into circulation as free antigen which can bind with MAbs. Thus extensive receptor shedding may accelerate clearance or decrease free MAb through competitive binding [12]. TMC is generally dependent on disease type and severity. Thus patients with more extensive disease and higher antigen burden will tend to have a higher fraction of MAb clearance through TMC. Even infliximab clearance, which does not exhibit TMC, has reported different clearances for different diseases which reflect the extent and degree of inflammation [13]. Faster MAb clearance in IBD is typically associated with low albumin which is also associated with more severe disease [2, 3]. Similarly diabetes is associated with faster MAb clearance. Diabetic comorbidity resulted in a 28% increase in ustekinumab clearance [14] which may be related to non-enzymatic glycation of proteins, which are cleared faster than non-glycated proteins [15].

Endogenous antibodies, MAbs, and albumin are protected from proteolysis by the Brambell

neonatal receptor (FcRn) [16], prolonging half-life [17]. FcRn receptors can be saturated at high IgG concentrations, resulting in shorter half-life [18]. Thus, albumin is often identified as being predictive of MAb clearance. In some disease states, such as multiple myeloma, high production of IgG M proteins results in a shortened half-life through FcRn saturation [19]. FcRn binding is species specific, so half-life generally increases as MAb structure becomes more human, with murine MAbs having a very short half-life (1-2 days); chimeric MAbs having a half-life of approximately 10-14 days, and fully human MAb having half-lives generally greater than 15 days [20]. Three additional Fc gamma receptors (FcγRI, FcγII and FcγIII) have been identified [21] which are expressed by macrophages, natural killer cells, B and T cells, and platelets. Fc gamma receptor binding elicits complement or antibody dependent cell cytotoxicity [22] which can form an additional route of clearance. MAb clearance through the reticuloendothelial system (RES) is partly regulated through interactions with FcγRs. The co-administration of methotrexate has been reported to reduce adalimumab clearance by 29-44% [23] in patients with rheumatoid arthritis. While methotrexate reduces immunogenicity-related clearance of adalimumab [24], it has been reported to reduce Fcγ receptor expression *in vitro* [25].

Regardless of extent of humanization, MAbs are exogenous proteins and all MAbs can induce ADA [26]. Numerous factors can impact ADA formation, including the formulation, its stability, extent of humanization, dose regimen, and treatment duration [27]. The intravenous (IV) route of administration is generally least likely to induce an ADA response, although this is not always true [28]. Subcutaneous administration is generally more immunogenic than IV. ADA occurs more frequently following administration of low doses than with high doses [29], and has been associated with intermittent exposure during clinical care [30]. ADAs can be neutralizing, in which MAb binding is impaired, or can increase MAb clearance [4]. Concomitant administration of immunosuppressants generally reduces the likelihood that a patient will develop ADA [31]. Concomitant administration of infliximab and methotrexate resulted in significantly lower ADA prevalence and generally higher serum infliximab concentrations, although without significantly impacting efficacy [32]. A report on Crohn's Disease (CD) patients who developed LOR to infliximab accompanied by ADA, showed the addition of an immunomodulator resulted in restoration of clinical response, decrease in ADA titers and increased infliximab trough concentrations [33].

Inflammatory diseases (ID) (e.g. Rheumatoid arthritis (RA), ankylosing spondylitis, inflammatory bowel disease (IBD), and psoriasis) are treated using "step-up" approaches, starting with chemical anti-inflammatory and immunomodulatory agents. Patients failing these therapies require treatment with monoclonal antibodies (MAbs) generally targeting tumor necrosis factor (TNF). While MAbs are effective treatments for ID, many patients lose response over time. In a retrospective assessment [34], discontinuation at 4 years for etanercept was 41%, infliximab was 46% and adalimumab was 52%. Approximately 20-30% of initially responding patients lose response during the first year of therapy [35] and subsequently approximately 10% lose response annually [36]. Psoriasis has similar failure rates.

Post-hoc analyses from pivotal trials in IBD suggested maintaining measurable serum infliximab trough concentrations during maintenance was associated with improved outcomes. [37, 38]. In a prospective study in patients with CD with secondary LOR to infliximab Steenholdt *et al.* [39] showed using an individualized therapeutic drug monitoring (TDM)-based dosing algorithm was cost-effective versus clinical symptom-driven dose escalations. A larger 1 year prospective trial [40] provided evidence that TDM-guided dosing

may preserve clinical response in patients with IBD after baseline adjustment of infliximab serum trough concentrations to 3 to 7 µg/ml. However, the utility of TDM for MABs has been questioned, partly because of lack of powered prospective studies using TDM-based dosing [41], together with a small prospective study (TAILORIX), investigating only dose increases in maintenance over 1 year, but not shortening dosing intervals, an important adjustment. The study design was insufficient to demonstrate the advantage (or lack) of TDM [42] but suggested no benefit. TDM utility for MABs has also been questioned due to slow assay turnaround, analytical deficiencies, assay differences, and difficulties with interpreting TDM [43]. These deficiencies are reasons that US insurance companies will generally not reimburse for MAB TDM [44]. The lack of reimbursement, together with the cost of the MAB assays (US\$250.00 to US\$2500.00) has compromised TDM applicability in the ID setting.

Identifying an individual's effective dose is neither intuitive nor static owing to flux in patient status and associated factors over time. This is particularly true for pediatric IBD using infliximab, which uses weight-based resulting in lower drug exposure in pediatrics [45]. Dashboard systems facilitate personalized dose adjustments using modeling, making better use of TDM [46]. A retrospective study using a prototype dashboard demonstrated quicker identification of individualized optimal dosage and identified LOR in advance of observed sub-therapeutic trough concentrations based on increasing individual clearance [47]. Another retrospective assessment of this dashboard found treatment recommendations were substantially different from standard of care, but feasible, and showed that patients recommended to have a dose adjustment had lower probability of clinical remission [48].

Loss of response to anti-TNF treatment is very common, and should be avoided if at all possible. Many retrospective studies have shown that LOR is often associated with low MAB serum concentrations. Reasons for insufficient exposure can vary a great deal and include lack of compliance, increasing disease activity and inflammatory burden and, importantly, formation of ADA. The presence of ADA is closely associated with relatively lower serum concentrations of drug. ADA formation can occur as early as 18 days after the initiation of treatment [49]. As a consequence it is of paramount importance to the clinician to have information on circulating concentrations of both MAB and ADA to allow for a comprehensive decision on dose adjustments or to recommend a switch to a different treatment, within or outside the therapeutic class.

The application of Bayesian dashboard systems to therapeutic MABs used to treat ID was selected owing to the current need to determine appropriate doses quickly to avoid intermittent exposure which can lead to ADA and LOR. In addition these agents are expensive and there is a strong desire to contain healthcare costs while improving patient outcomes. Dashboard systems make the transition from passive TDM to proactive therapeutic management taking into account between subject differences in both pharmacokinetics and pharmacodynamics, and as such are expected to show clinical benefit. A prototype system is currently undergoing testing in a clinical trials to determine its usefulness, and if useful, will result in the system being expanded to include other MABs used to treat IBD. Application in other therapeutic areas including RA and psoriasis will also need testing although the failure rate for MABs in RA is comparable to that in IBD and TDM is already being adopted by rheumatologists.

References

1. Waldmann TA, Strober W. Metabolism of immunoglobulins. *Prog. Allergy*. 1969; 13:1–110.
2. Xu Z, Mould DR, Hu C, Ford J, Keen M, Davis HM, Zhou H. “A Population-Based Pharmacokinetic Pooled Analysis of Infliximab in Pediatrics” ACCP National Meeting 2012 San Diego CA
3. Fasanmade AA, Adedokun OJ, Blank M, et al. Pharmacokinetic properties of infliximab in children and adults with Crohn's disease: a retrospective analysis of data from 2 phase III clinical trials. *Clin Ther*. 2011;33(7):946–64.
4. Mould DR. “Using Pharmacometrics in the Development of Biological Therapeutic Biological Agents” In *Pharmacometrics: The Science of Quantitative Pharmacology* Editors: E. Ette and P Williams, Chapter 41 John Wiley and Sons Hoboken, NJ 2007
5. Roberts BV, Susano I, Gipson DS, Trachtman H, Joy MS. Contribution of renal and non-renal clearance on increased total clearance of adalimumab in glomerular disease. *J Clin Pharmacol*. 2013;53(9):919-24.
6. Beeken WL Busch HJ, Sylwester DL. Intestinal protein loss in Crohn's disease. *Gastroenterology* 1972 ;62(2):207-15.
7. Kapel N, Meillet D, Favennec L, Magne D, Raichvarg D, Gobert JG. Evaluation of intestinal clearance and faecal excretion of alpha 1-antitrypsin and immunoglobulins during Crohn's disease and ulcerative colitis. *Eur J Clin Chem Clin Biochem*. 1992;30:197-202
8. Brandse JF, Wildenberg M, de Bruyn JR, et al. Fecal loss of infliximab as a cause of lack of response in severe inflammatory bowel disease. *Gastroenterology* 2013;144(5):S-7763
9. Levy J, Barnett EV, MacDonald NS, Klinenberg JR. Altered immunoglobulin metabolism in systemic lupus erythematosus and rheumatoid arthritis. *J Clin Invest*. 1970;49(4):708-15.
10. Ordás I, Mould DR, Feagan BG, Sandborn WJ. “Monoclonal Antibodies in Inflammatory Bowel Disease: Pharmacokinetic Based Dosing Paradigms.” *Clin. Pharmacol. Ther*. 2012; 91(4):635-46
11. Mould DR, Baumann A, Kuhlmann J, et al. Population Pharmacokinetics pharmacodynamics of alemtuzumab (Campath) in patients with chronic lymphocytic leukemia. *Br J Clin Pharmacol* 2007; 64 (3): 278-91
12. Mould DR, Green B. *Pharmacokinetics and Pharmacodynamics of Monoclonal Antibodies: Concepts and Lessons for Drug Development*. 2010; *BioDrugs* 24(1):23-39.
13. Mould DR Commentary: Why Therapeutic Drug Monitoring is Needed for Monoclonal Antibodies and How Do We Implement This? *Clin Pharmacol Ther*. 2016;99(4):351-4
14. Zhu Y, Hu C, Lu M, Liao S, Marini JC, Yohrling J, Yeilding N, Davis HM, Zhou H. Population pharmacokinetic modeling of ustekinumab, a human monoclonal antibody targeting IL-12/23p40, in patients with moderate to severe plaque psoriasis. *J Clin Pharmacol*. 2009;49(2):162-75.
15. Kaneshige H. Nonenzymatic glycosylation of serum IgG and its effect on antibody activity in patients with diabetes mellitus. *Diabetes*. 1987;36(7):822-8.
16. Brambell FW, Hemmings WA, Morris IG. A Theoretical Model of Gamma-Globulin Catabolism. *Nature*. 1964; 203: 1352-4

17. Telleman P, Junghans RP. The role of the Brambell receptor (FcRB) in liver: protection of endocytosed immunoglobulin G (IgG) from catabolism in hepatocytes rather than transport of IgG to bile. *Immunology*. 2000; 100(2): 245-51.
18. Morell A, Terry WD, Waldmann TA. Metabolic properties of IgG subclasses in man. *J. Clin. Invest.* 1970; 49(4): 673-80
19. Jacobs J, Mould DR. The Role of FcRn in the Pharmacokinetics of Biologics in Patients With Multiple Myeloma. *Clin Pharmacol Ther.* 2017 Feb 10. [Epub]
20. Ternant D PG. Pharmacokinetics and concentration-effect relationships of therapeutic monoclonal antibodies and fusion proteins. *Expert. Opin. Biol. Ther.* 2005; 5((Suppl 1)): S37-47.
21. Cohen-Solal JF, Cassard L, Fridman WH, Sautes-Fridman C. Fc gamma receptors. *Immunol. Lett.* 2004; 92(3): 199-205
22. Reddy MP, Kinney CA, Chaikin MA, et al. Elimination of Fc receptor dependent effector functions of a modified IgG4 monoclonal antibody to human CD4. *J Immunol* 2000; 164 (4): 1925-33
23. Seitz K, Zhou H. Pharmacokinetic drug-drug interaction potentials for therapeutic monoclonal antibodies: reality check. *J Clin Pharmacol.* 2007;47(9):1104-18.
24. Bartelds GM, Wijbrandts CA, Nurmohamed MT, et al. Clinical response to adalimumab: relationship to anti-adalimumab antibodies and serum adalimumab concentrations in rheumatoid arthritis. *Annals of the rheumatic diseases.* 2007; 66(7):921-6.
25. Wijngaarden S, van Roon JA, van de Winkel JG, Bijlsma JW, Lafeber FP. Down-regulation of activating Fcγ receptors on monocytes of patients with rheumatoid arthritis upon methotrexate treatment. *Rheumatology.* 2005; 44(6):729-34.
26. Weiner LM. Monoclonal antibody therapy of cancer. *Semin Oncol* 1999; 26: 43-51
27. Schellekens H. Bioequivalence and the immunogenicity of biopharmaceuticals. *Nat Rev Drug Discov.* 2002;1(6):457-62.
28. Kijanka G, Jiskoot W, Schellekens H, Brinks V. Effect of treatment regimen on the immunogenicity of human interferon Beta in immune tolerant mice. *Pharm Res.* 2013;30(6):1553-60.
29. Stephens S, Emtage S, Vetterlein O, Chaplin L, Bebbington C, Nesbitt A, Sopwith M, Athwal D, Novak C, Bodmer M. Comprehensive pharmacokinetics of a humanized antibody and analysis of residual anti-idiotypic responses. *Immunology.* 1995; 85(4):668-74.
30. Brandse JF, Mould DR, Ashruf YK, Smeekes OS, Kuin S, van den Brink GR, D'Haens GR. Factors that increase clearance of infliximab: relationship between insufficient infliximab exposure and antidrug antibody formation in IBD, a pharmacokinetic study. *Inflam Bowel Dis.* 2017 ;23(4):650-660
31. Colombel JF, Sandborn WJ, Reinisch W, et al. Infliximab, azathioprine, or combination therapy for Crohn's disease. *N Engl J Med* 2010;362:1383-1395.
32. Feagan BG, McDonald JW, Panaccione R, et al. Methotrexate in combination with infliximab is no more effective than infliximab alone in patients with Crohn's disease. *Gastroenterology* 2014;146:681-688.e1.
33. Ben-Horin S, Waterman M, Kopylov U, et al. Addition of an immunomodulator to infliximab therapy eliminates antidrug antibodies in serum and restores clinical response of patients with inflammatory bowel disease. *Clin Gastroenterol Hepatol* 2013;11:444-447.

34. Souto A, Maneiro JR, Gómez-Reino JJ. Rate of discontinuation and drug survival of biologic therapies in rheumatoid arthritis: a systematic review and meta-analysis of drug registries and health care databases. *Rheumatology (Oxford)*. 2016;55(3):523-34
35. Peyrin-Biroulet L, Deltenre P, de Suray N, Branche J, Sandborn WJ, Colombel JF. Efficacy and safety of tumor necrosis factor antagonists in Crohn's disease: meta-analysis of placebo-controlled trials. *ClinGastroenterolHepatol*. 2008;6(6):644-53.
36. Gisbert JP, Panes J. Loss of response and requirement of infliximab dose intensification in Crohn's disease: a review. *Am J Gastroenterol* 2009;104:760-7.
37. Fasanmade AA, Adedokun OJ, Ford J, et al. Population pharmacokinetic analysis of infliximab in patients with ulcerative colitis. *Eur J ClinPharmacol* 2009;65:1211-28.
38. Reinisch W, Colombel JF, Sandborn WJ, Mantzaris GJ, Kornbluth A, Adedokun OJ, Miller M, Tang KL, Rutgeerts P, Cornillie F. Factors associated with short- and long-term outcomes of therapy for Crohn's disease. *ClinGastroenterolHepatol*. 2015 Mar;13(3):539-547..
39. Steenholdt C, Brynskov J, Thomsen OO, et al. Individualised therapy is more cost-effective than dose intensification in patients with Crohn's disease who lose response to anti-TNF treatment: a randomised, controlled trial. *Gut* 2014;63:919-27
40. Vande Castele N, Gils A. Preemptive Dose Optimization Using Therapeutic Drug Monitoring for Biologic Therapy of Crohn's Disease: Avoiding Failure While Lowering Costs? *Dig Dis Sci*. 2015 Apr 28.
41. Mould DR. Commentary: Why Therapeutic Drug Monitoring is Needed for Monoclonal Antibodies and How Do We Implement This? *Clin Pharmacol Ther*. 2016;99(4):351-4
42. D'Haens GR, Vermeire S, Lambrecht G, Baert FJ, Bossuyt P, Nachury M, Buisson A, Bouhnik Y, Filippi J, van der Woude CJ, van Hootegem P, Moreau J, Louis E, Franchimont D, De vos M, Mana F, Peyrin-Biroulet L, Brix H, Allez M, Caenepeel P, Aubourg A, Oldenburg B, Pierik M, Chevret S, Gils A, Laharie D. 692 Drug-Level Based Dosing Versus Symptom-Based Dose Adaptation in Patients With Crohn's Disease: A Prospective, Randomized Multicenter Study (TAILORIX). *Gastroent*. 2016 150(4):S143Stroh M, Lum BL. Commentary: Should Therapeutic Drug Monitoring for Monoclonal Antibodies Remain the Exception or Become the Norm? *Clin Pharmacol Ther*. 2016; 100(6): 215–217
43. Stroh M, Lum BL. Commentary: Should Therapeutic Drug Monitoring for Monoclonal Antibodies Remain the Exception or Become the Norm? *Clin Pharmacol Ther*. 2016; 100(6): 215–217
44. http://www.aetna.com/cpb/medical/data/300_399/0341.html
45. Zhao Q, Tensfeldt TG, Chandra R et al. Population Pharmacokinetics of Azithromycin and Chloroquine in Healthy Adults and Pediatric Malaria Subjects Following Oral Administration of Fixed Dose Azithromycin and Chloroquine Combination Tablets. *Malar J*. 2014;13:36
46. Mould DR, Dubinsky MC. Dashboard Systems: Pharmacokinetic / Pharmacodynamic Mediated Dose Optimization. *J Clin Pharmacol*. 2015;55 Suppl 3:S51-9
47. Mould DR, Moyer B, Amur S *et al*. “The Impact of New Technologies on the Science of Clinical Care and Drug Development”. *AAPS Magazine* December 2013 67.

48. Dubinsky MC, Phan BL, Singh N, Rabizadeh S, Mould DR. Pharmacokinetic dashboard recommended dosing is different than standard of care dosing in infliximab treated pediatric IBD patients. *AAPS J.* 2017;19(1):215-222
49. Brandse JF, Mathôt RA, van der Kleij D, Rispens T, Ashruf Y, Jansen JM, Rietdijk S, Löwenberg M, Ponsioen CY, Singh S, van den Brink GR, D'Haens GR. Pharmacokinetic Features and Presence of Antidrug Antibodies Associate With Response to Infliximab Induction Therapy in Patients With Moderate to Severe Ulcerative Colitis. *Clin Gastroenterol Hepatol.* 2016 ;14(2):251-8
- 50.